

Next-Generation Morphology (NGM) Enables Advanced Digital Analysis of Myelodysplastic Syndromes in Peripheral Blood Smears

A multilayer morphometric approach delivers high-resolution Full-Field imaging and quantitative insights into myelodysplasia using the Scopio Labs X100 digital system. ^{1 a p1, a-c p3 Katz}



Background

Morphological analysis has not kept pace with molecular and flow cytometry advancements using AI tools, remaining largely manual, low in sensitivity, and constrained by technological and theoretical limitations. ^{1 b p1 Katz}

A fundamental trade-off between field of view and resolution restricts high-magnification imaging ($\times 100$) to only 100–200 leukocytes, limiting analytical sensitivity to around 10^{-2} . ^{1 c p1, d p3 Katz}

Achieving high-resolution, large-scale (“big data”) imaging within practical timeframes remains challenging. ^{1 d p1 Katz}

~80% of practitioners continue to rely on manual microscopy for peripheral blood smear (PBS) analysis. ^{1 e p3 Katz}

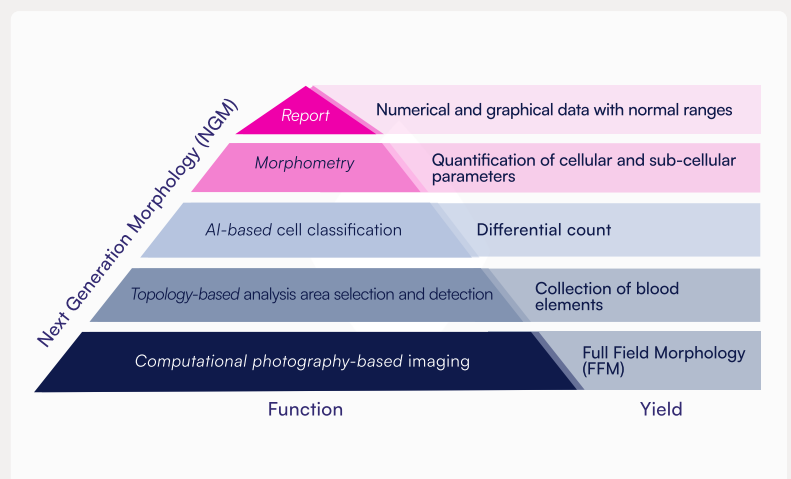
A prospective study evaluated next-generation morphology (NGM). The Scopio Labs X100 platform was used to test NGM performance by analyzing peripheral blood smears (PBS) from myelodysplastic syndrome (MDS) patients and age-matched controls. ^{1 e p1, c p3 Katz}



“NGM directly addresses the gaps in high-resolution digitization detailed previously, and unlock the diagnostic and research potentials of morphology in hematology.” ^{1 f p3 Katz}

Methodology

- Stained PBS utilizing a May–Grunwald–Giemsa protocol were analyzed from 37 myelodysplastic syndrome (MDS) patients and 30 age-matched controls. ^{1 e p1, a p11 Katz}
- A low-magnification objective coupled with an illumination array captured multiple large field-of-view (FOV) low-resolution images. Each image was taken under different wavelengths and illumination angles, capturing unique data patterns from the sample. ^{1 g p3 Katz} A high-resolution image was computationally reconstructed from these multiple low-resolution inputs using physical imaging models. ^{1 h p3 Katz}
- AI and machine learning (ML) algorithms were integrated with traditional computer vision for cell detection and classification: ^{1 i p3 Katz}



Neutrophils granulation index:

Using a simple dimensionality reduction-based algorithm, a single grading scale without units (granulation index [GI]) was created from the parameters. A dataset of representative samples for each neutrophil population (hypogranulation, normal, toxic granulation, five of each) was used to cover the entire granulation spectrum in the scale creation. ^{1 a p4 Katz}

RBC morphometry:

Once RBCs are properly segmented, geometric-based features can be extracted with image processing algorithms. The cell size is derived from the segment size and is quantified in area units (μm^2). The deformation of the cell contour is derived from the normalized distribution of the contour distances from the cell center point and is quantified in length units (μm). ^{1 b p4 Katz}

IPSS-R MDS patient's stratification and statistics:

The Revised International Prognostic Scoring System (IPSS-R) assessed MDS risk using CBC parameters (hemoglobin, ANC, platelets), cytogenetics, and blast percentage, with scores >2.5 classified as high-risk. Demographic and laboratory data were analyzed using univariate and multivariate Cox regression, adjusted for age, gender, and morphometric variables, to identify independent predictors of MDS risk ($p < 0.05$ considered significant). ^{1 a p5 Katz}

1 Clinical Site ^{1 a p11 Katz}

Tel Aviv Sourasky Medical Center ^{1 j p3 Katz}

What Did They Evaluate?

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- Utilization of next-generation morphology (NGM) for precise quantitative analysis of myelodysplasia in PBS ^{1 k p3 Katz}

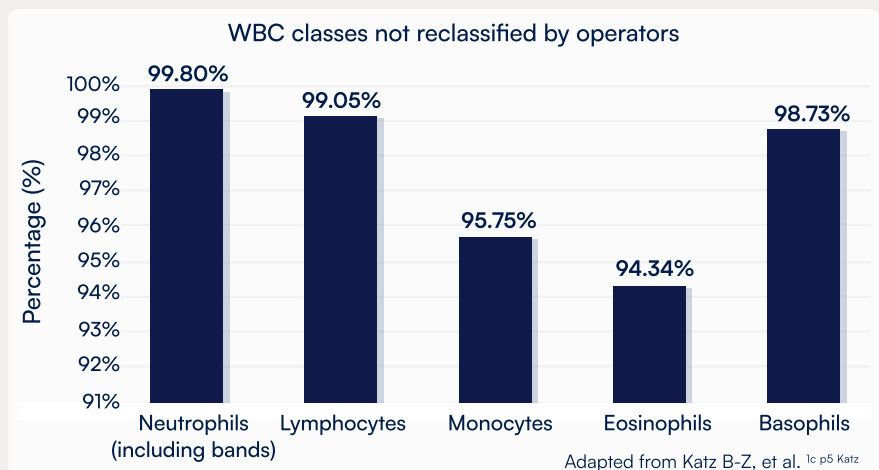
Results

>94% of the WBCs belonging to the five major WBC classes were correctly classified by the system ^{1 b p5 Katz}

274,856 cells analyzed ^{1 c p5 Katz}
Representing 1374 PBS

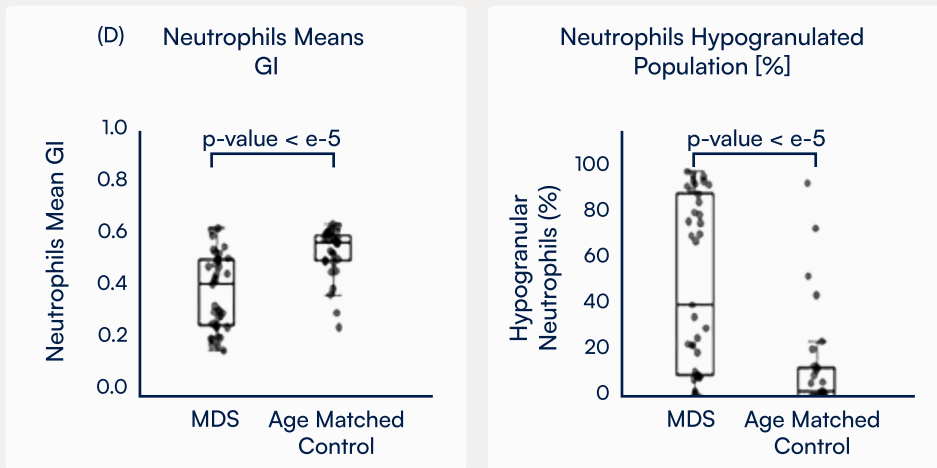
These cells were rarely reclassified by human operators—accuracy was highest for neutrophils (99.8%) and basophils (98.73%). ^{1 c p5 Katz}

- Only 0.41% of cells were initially unclassified; operators most often reassigned these as neutrophils (26.68%) or blasts (12.85%), while 39.89% remained unclassified. ^{1 e p5 Katz}
- Blasts were correctly identified 79.91% of the time, with misclassifications mainly as lymphocytes or monocytes. ^{1 f p5 Katz}



Significant reduction in mean GI in MDS

- Due to the high accuracy of AI-based neutrophil classification, neutrophils were evaluated automatically - without human review - in 37 MDS samples and 30 age-matched control samples. ^{1 g p5, a p6 Katz}

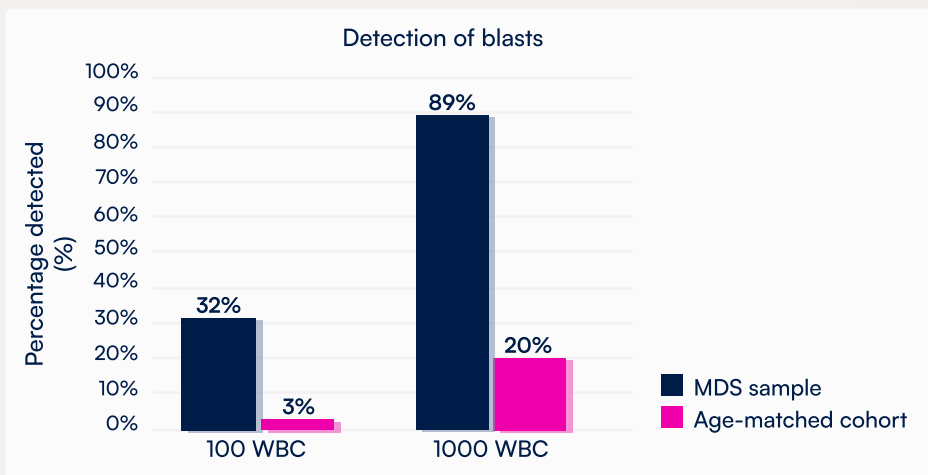


Adapted from Katz B-Z, et al. ^{1 a p8 Katz}

- Mean GI significantly reduced in MDS vs controls (0.38 ± 0.14 vs 0.53 ± 0.10 , $p < 10^{-5}$). ^{1 a p7 Katz}
- Hypogranulated neutrophils increased (49.6% vs 13.2% , $p < 10^{-5}$). ^{1 b p7 Katz}

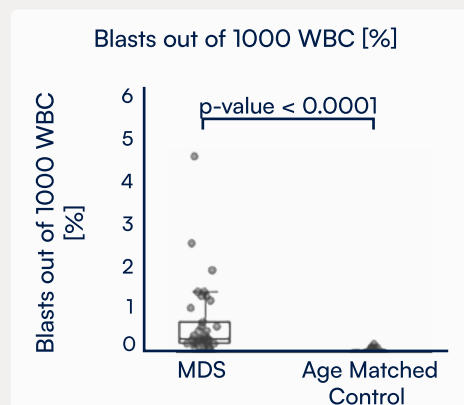
Enhanced detection sensitivity for blast enumeration

- Blasts percentage per 100 (according to the current practice) or 1000 (now provided by the FFM) WBC counts/sample in 37 MDS and 30 age-matched control samples. ^{1 b p8 Katz}
- A highly significant difference ($p < 10^{-9}$) in the detection of blasts was noted in 1000 WBC counts per sample. ^{1 c p8 Katz}



Adapted from Katz B-Z, et al. ^{1 c,d p8 Katz}

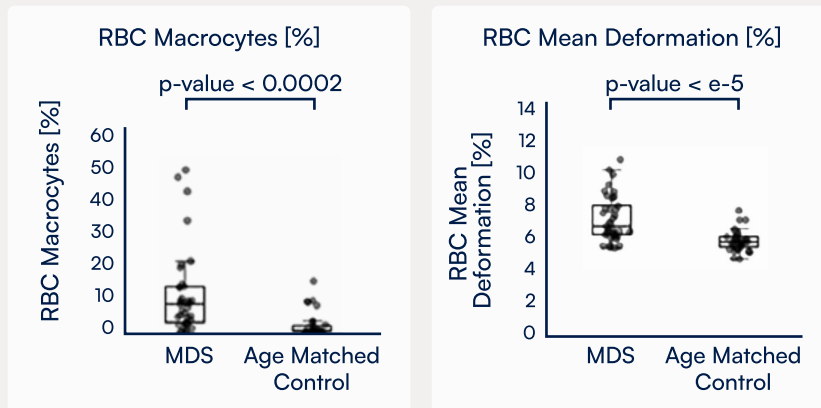
- The percentage of blasts per 1,000 WBCs in MDS samples was $0.65 \pm 0.87\%$ (range: 0—4.50%), which was significantly higher ($p < 0.0001$) than in age-matched controls, who showed $0.02 \pm 0.05\%$ blasts (range: 0—0.20%). ^{1 e p8 Katz}



Adapted from Katz B-Z, et al. ^{1 a p9 Katz}

Detection of RBC morphometry

- At least 152,310 red blood cells (RBCs) were analyzed per peripheral blood smear (PBS) to ensure a 95% confidence interval for detecting RBC abnormalities occurring at a 1% frequency.^{1 f p8 Katz}



Adapted from Katz B-Z, et al. ^{1 a p10 Katz}

- Macrocytosis was significantly higher in MDS samples ($11.44 \pm 11.89\%$, range 0.64–46.83%) compared with age-matched controls ($2.85 \pm 3.72\%$, range 0.34–15.30%; $p < 0.0002$).^{1 g p8 Katz}
- MDS samples also had a significantly higher proportion of deformed RBCs ($7.24 \pm 1.42\%$, range 5.4–10.82%) than controls ($5.86 \pm 0.67\%$, range 4.75–7.74%; $p < 10^{-5}$).^{1 h p8 Katz}

Conclusions

The Next-Generation Morphology (NGM) platform using Scopio Labs' X100 scanner and application enables:

- High-resolution, large-field imaging digital PBS analysis.^{1 a p1 Katz}
- Sensitive detection of morphologic abnormalities in MDS.^{1 g p1 Katz}
- Objective, quantitative morphometry supporting risk stratification.^{1 a p1 Katz}
- Opportunities for remote diagnostics and research into hematologic dysplasia.^{1 b p9 Katz}

[Read the full publication](#)

1. Katz B-Z, Moshe Y, Bensity D, Luttwak E, Brazilai M, Oster HS, et al. Next-generation morphology, a novel multilayer morphometric digital analysis, reveals both the basic topology and new trends of myelodysplasia of peripheral blood specimens. *Br J Haematol.* 2025;00:1–12. doi:10.1111/bjh.70110